

Mini-cutting of *Plathymenia reticulata* benth. wit mini-stumps conducted in suspended seedbed and tubes

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SILVICULTURE

ABSTRACT

Background: Despite the importance of *Plathymenia reticulata* for forestry and ecological purposes, a protocol for the vegetative propagation of the species remains unclear mainly due to the low adventitious rooting of the propagules of the species. In this sense, this work aimed to evaluate the rooting of mini-cuttings of juvenile materials of *P. reticulata* and our hypothesis: (1) maintaining propagules under mist chamber for different periods affects the rooting process; (2) the mini-stump management system due to the restriction of the root system affects the production of mini-cuttings and the quality of clonal seedlings.

Results: It was not necessary for more than 30 days in a mist chamber to induce rooting. The highest percentage of rooting was obtained 50 days after staking. Mini-gardens set up in suspended seedbed and tubes (280 cm³) had the average mini-cuttings productivity of 4.32 and 2.06, respectively, over 270 days of exploration (monthly collections). At 120 days after staking, there was no difference in the survival and in the final quality of clonal seedlings produced. However, clonal seedlings from mini-garden in tubes had higher height, leaf area, and number of first-order roots. Regardless of the mini-garden management, the clonal seedling production index was 50%.

Conclusion: We can conclude that *P. reticulata* seedling production via mini-cutting technique is possible.

Keywords: Adventitious rooting, Cloning, Native forest species, Vegetative propagation

HIGHLIGHTS

It is not necessary more that 30 days in a mist chamber to *P. reticulata* rooting.
The suspended seedbed mini-garden showed the highest production of mini-cuttings.
The root restriction of mini-stumps didn't affect the quality of the clonal seedlings.
The clonal seedling production rate for both types of mini-gardens was 50%.

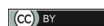
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INTRODUCTION

Plathymenia reticulata Benth. is a tropical forest species of the Fabaceae family. The species is spread throughout the Brazilian states, occurring in the Amazon, Cerrado, Caatinga, and Atlantic Forest biomes (Flora do Brasil, 2020; Freire *et al.*, 2017). The species has presented high survival rates and growth after outplanting (Barroso *et al.*, 2018), and its timber is valuable due to its high quality, ease of handling, and resistance to xylophagous organisms, qualities that ensure the importance of this species (Lorenzi, 2002; Carvalho, 2009).

P. reticulata seedling production is mostly carried out using seeds. However, Pessanha *et al.* (2018) reported that matrices have shown irregular behavior in seed production. Another drawback to the use of sexual propagation involves the implantation of heterogeneous stands, an outcome that increases costs related to management practices and harvesting. In this sense, the vegetative propagation of the species has the advantage of creating homogenous plantations.

Among the vegetative propagation techniques applied to forest species, mini-cutting has been used on a commercial scale for the production of *Eucalyptus* spp. and *Toona ciliata* seedlings (Bela Vista Florestal, 2020). However, the results of various studies indicate the technique is feasible for other species, including *Cedrela fissilis* (Xavier *et al.*, 2003), *Ilex paraguariensis* (Wendling *et al.*, 2007), *Erythrina falcata* (Cunha *et al.*, 2008), *Calophyllum brasiliense* (Kalil Filho *et al.*, 2010; Silva *et al.*, 2010; Santos, 2013; Ciriello and Mori, 2015), *Cariniana legalis* (Gatti *et al.*, 2011), *Anadenanthera colubrina* (Dias *et al.*, 2012), *Handroanthus heptaphyllus* (Oliveira *et al.*, 2015; Oliveira *et al.*, 2016), *Lecythis pisonis* (Santana, 2017), *Inga edulis* (Berude *et al.*, 2019), and *Cabralea canjerana* (Burin *et al.*, 2018a and 2018b).

Several factors can drive the rooting process (jointly or independently) such as humidity, temperature, light, substrate, genotype, physiological and nutritional conditions, age of the mother plant, size and type of propagule, presence of leaves, and collection time (Xavier *et al.*, 2003b). In this context, the period during which a propagule remains in each phase (e.g. mist chamber and shade house) may influence root survival rates (Ferreira *et al.*, 2004).

The growth conditions to which the mini-stumps are submitted may also influence sprouting capacity and root quality (Wendling *et al.*, 2005; Souza Junior, 2007; Cunha *et al.*, 2008; Silva *et al.*, 2012; Souza *et al.*, 2014b). For *P. reticulata*, the low adventitious rooting rates reported by Neubert *et al.* (2017), Pessanha *et al.* (2018), and Rodrigues (2018) may have discouraged seedling production with this technique. Studies investigating stepwise and management of the process are needed in order to enhance understanding of the main factors driving *P. reticulata* rooting.

Thus, this work aimed to evaluate whether increasing the time *P. reticulata* mini-cuttings remain in the rooting sector (mist chamber) results in improved rooting and to help establish the ideal time mini-cuttings should remain in this sector. It also aimed to verify if root system restriction influences the quality of the clonal seedlings.

MATERIAL AND METHODS

The study was carried out in a greenhouse covered with plastic (150 μm) at the Universidade Federal do Norte Fluminense Darcy Ribeiro - UENF, within the municipal limits of Campos dos Goytacazes, RJ, Brazil (21° 19' 23" S and 41° 19' 41" W). The experimental scheme is presented in Figure 1. In the rooting sector, using data logger model RHT10 (Extech Instruments), with readings programmed every hour, the average maximum and minimum temperature were 26.9 and 19.4°C, respectively, while the average maximum and minimum humidity were 87.3 and 52.6%, respectively.

Experiment 1 - Time in mist chamber

The mini-cuttings were collected from a seminal mini-garden, and each mini-cutting was 6 cm in length. One pair of leaves was retained on each mini-cutting with the leaflets trimmed 50% to reduce the umbrella effect, which prevents water from reaching the substrate. Then 430 mini-cuttings were staked in tubes of 120 cm³ filled with substrate composed of pine bark, coconut fiber, peat fiber, and vermiculite. We added 8 g of slow-release fertilizer (N P K - 15-9-12) per kg⁻¹ of the substrate.

The mini-cuttings were sent to an intermittent mist chamber (rooting sector) covered with 150 μm polypropylene plastic, 30% shading cloth (Sombrite), and foil shading (Aluminet - 30% shading). The mist flow was 3 mL min⁻¹m⁻² and the nebulization frequency approximately every 20 minutes for 30 seconds. After 30 days in the rooting sector the survival rate was 66.5% (286 mini-cuttings). The dead mini-cuttings were discarded and 40 of the surviving cuttings were randomly selected (10 mini-cuttings per plot) for rooting assessment.

Then, 50% (120 mini-cuttings) of the surviving mini-cuttings were then moved to a greenhouse with a plastic cover (150 μm agricultural film) and 30% shading cloth (Sombrite) and automatic sprinkler with irrigation three times a day for approximately 8-10 minutes. In the greenhouse the mini-cuttings were arranged in a completely randomized design with 12 plots and 10 mini-cuttings per plot with 4 repetitions per treatment. The mini-cuttings that remained in the rooting sector were subjected to the same experimental design and sampling procedure. The rooting assessments, for both environments, were carried out every ten days up to 60 days after staking and 4 plots (40 mini-cuttings) were randomly removed at each time.

For each rooting assessment, after washing the root system under tap water we evaluated the presence of calluses and the number and length of first order roots (NFOR and LFOR). We considered rooted mini-cuttings those possessing at least one root greater than 0.2 mm. The samples were subsequently oven-dried at 65°C \pm 2 for 72 h and weighed to determine the dry mass of the roots (DMR).

For data analysis, a completely randomized design under a 2 x 3 factorial scheme was considered. The environmental conditions (mist chamber and greenhouse) were considered the main factor and the subplot factor was the assessment time (40, 50, and 60 days). The mini-

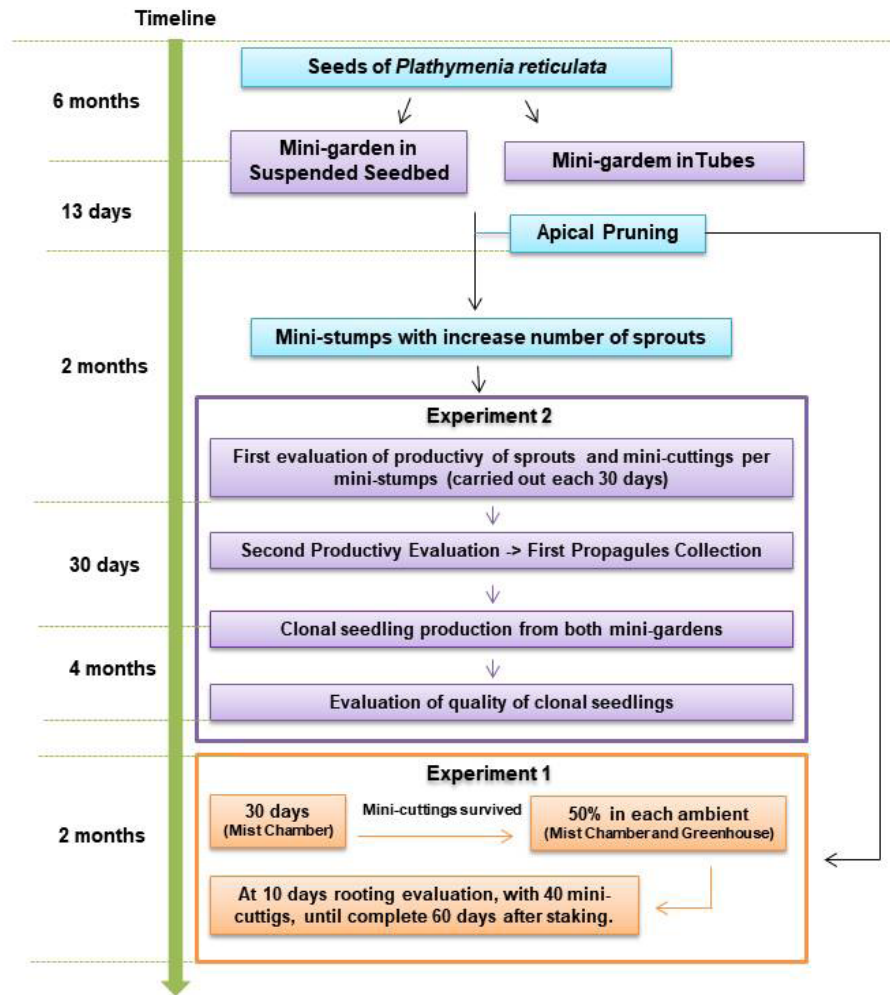


Fig. 1 Flowchart of experimental procedure for assessing the vegetative propagation of *Plathymenia reticulata* Benth. with the mini-cutting technique.

cuttings removed 30 days after staking were considered an additional treatment with 4 replicates and 10 mini-cuttings per plot. Then, two-way ANOVA and Tukey tests ($p < 0.05$) were performed to test for the individual and interactive effects of the treatments (main factor and subplot factor) – agricolae package, Mendiburu, (2019) through R environment (R Core Team, 2019).

Experiment 2 - Mini-garden management and clonal seedling production

Two mini-gardens treatments were established: (1) 36 mini-stumps in suspended seedbeds and (2) 36 mini-stumps in tubes (180 cm³). The seedbed sizes were 113 x 105 x 28 cm (length, width, and height, respectively), and the mini-stumps were spaced at 17 x 18 cm. The substrate for the seedbed was composed of an 8 cm layer of sand below a 20 cm layer containing a mixture of pine bark substrate, coconut fiber, and 2:1:1 organic fertilizer along with 600 g of simple superphosphate. Then the seedlings of both treatments had their apices pruned 10 cm from the base to form the mini-stumps.

Nitrogen fertilization was performed twice in the mini-gardens, 120 and 210 days after mini-stump formation. The solution was prepared according to Gonçalves and Benedetti (2005) with 12 g.L⁻¹ of (NH₄)₂SO₄, 10 mL per mini-stump. Mini-garden productivity for each treatment was measured monthly up to 270 days after mini-stump establishment, assessing the number of sprouts and mini-cuttings produced per mini-stump.

In the second productivity assessment, 8 cm long mini-cuttings containing a pair of leaves with the leaflets trimmed by 50% were detached. The mini-cuttings were staked in 180 cm³ tubes filled with pine bark substrate, coconut fiber, peat fiber, and vermiculite and slow release fertilizer (N P K – 15-9-12, with 8 g applied per Kg⁻¹ of the substrate). Each mini-garden treatment had 8 replicates with 18 plants in a completely randomized design. The trays with the mini-cuttings were sent to the rooting sector, where they remained for 30 days before being sent to the greenhouse.

$$DQI = \frac{TDM}{\left(\frac{H}{D}\right) + \left(\frac{DMAP}{DMSR}\right)} \tag{1}$$

The surviving clonal seedlings were evaluated for height (H) and stem diameter (D) 120 days after staking. Next, we measured leaf area using a bench area meter (LI – 3000 LI-COR Inc) and the samples were oven-dried at $65^{\circ}\text{C} \pm 2$ for 72 h to determine dry shoot mass (DMS). The root systems were then washed under tap water and callus presence, number of first order roots (NFOR), number of first order roots with branches (NRB), and the length of first order roots (LFOR) were assessed.

The total length of the root system (TLR), root diameter (DR), and volume (VR) were also measured with Winrhizo software (Regent Instruments Inc., Quebec, Canada, 2000). Finally the root samples were oven-dried at $65^{\circ}\text{C} \pm 2$ for 72 h to determine the dry mass of the root system (DMRS). We calculated the Dickson seedling quality index as described by Dickson et al. (1960) with the following equation: where TDM = total dry mass (g), DMS = shoot dry mass; DMSR = dry mass of root system (g); H = height (cm); D = stem diameter (mm).

Due to the high mini-cutting mortality during the experiment, the initial experimental design was modified. Thus, the surviving clonal seedlings from each mini-garden treatment were randomly redistributed for data analysis purposes with each treatment having 6 replicates with 12 plants in a completely randomized design.

Before proceeding to statistical analyses, quantitative data were converted into $\sqrt{x + 1}$ and the percentage data into arcsine \sqrt{x} (Zimmermann, 2004). The other data were submitted to diagnostic plots (Residuals fitted, Normal, and Leverage - base package, R Core Team, 2019) and the maximum likelihood function test (Box-Cox) – MASS package, Venables and Ripley, (2002) and the data transformed in Log ($x + 1$). Next, one-way ANOVA and Tukey tests ($p < 0.05$) were performed for treatment comparison - agricolae package, Mendiburu, (2019). The number of mini-cuttings and shoots and the number of accumulated mini-cuttings were submitted to descriptive analyses by confidence interval.

RESULTS

Experiment 1 - Time under mist chamber

The survival rate of mini-cuttings was 67% after 30 days in the rooting sector under intermittent misting. In the subsequent assessments of the remaining clonal seedlings, arranged in the greenhouse, the survival rate was 70%.

The highest percentage of *P. reticulata* rooting was observed 50 days after staking, and the environmental factor did not affect the rooting of mini-cuttings (Tab.1). In addition, treatments showed an average of 44% of mini-cuttings with callus formation, regardless of root emission.

Tab. 1 Mini-cutting rooting of *Plathymeria reticulata* Benth. at different times in the mist chamber and greenhouse. N = 40.

Time* (days)	Rooting (%)	
	Mist chamber	Greenhouse
40	0 bA	3 bA
50	10 aA	20 aA
60	15 abA	8 abA

*Days after staking. The means with the same lowercase letter in the line and the same uppercase letter in the column did not differ, according to the Tukey test, with a 5% significance.

Experiment 2 - Mini-garden management and clonal seedling production

The consecutive mini-cutting collections did not significantly affect the survival of the mini-stumps in both mini-garden management schemes. The averages for the survival of mini-stumps in suspended seedbeds and tubes was 83.33% and 100%, respectively, 270 days into the mini-gardens.

Mini-stumps in both types of mini-garden managements showed the ability and capacity to produce new shoots after each new sprout collection. However, there was a difference in the productivity of sprouts and mini-cuttings per mini-stump, as the mini-stumps established in tubes showed a higher number of sprouts. In contrast, the production of sprouts was the same in both mini-garden systems at 120, 150, and 270 days after the formation of mini-stumps (Fig.2 A and B).

The suspended seedbed mini-garden had the highest production of mini-cuttings at the end of 270 days after the formation of mini-stumps (Fig. 3).

Regardless of the mini-garden management technique, the survival rate was 50% considering the initial number of mini-cuttings staked (144 mini-cuttings per treatment). Although there was no difference in the production of dry biomass, clonal seedlings produced by mini-cuttings from the tube mini-garden possessed higher heights (H) and leaf area (LA) (Tab. 2).

Regarding the root system, there was a difference in the number of first order roots (NFOR) and the number of first order roots with branches (NRB) between the two mini-garden systems. Greater height measurements resulted with mini-cuttings from mini-stumps established in tubes (Tab. 2). The Dickson Quality Index values did not differ between the types of mini-gardens.

DISCUSSION

Experiment 1 - Time under mist chamber

Low rooting performance of mini-cuttings may be linked to physiological conditions and environmental factors such as temperature and humidity in the rooting sector (Xavier et al., 2013). However, during the present experiment, the temperature and humidity conditions of the mist chamber were conducive to rooting.

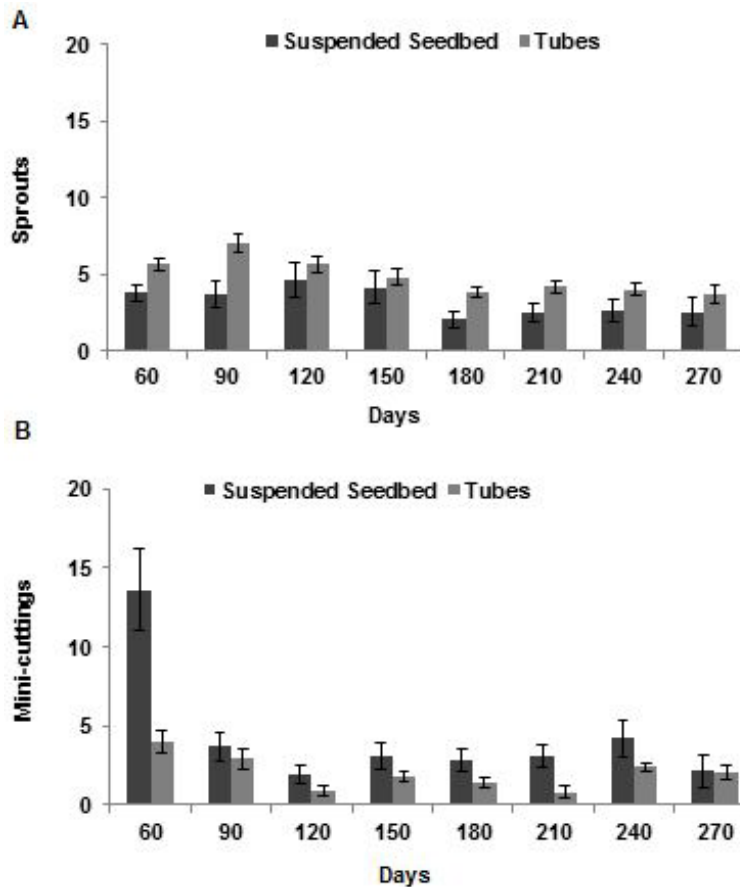


Fig. 2 Production of sprouts per mini-stump (A) and mini-cutting productivity per mini-stump (B) of *Plathymeria reticulata* Benth. carried out in two mini-gardens systems (suspended seedbed and tubes) for 270 days after the formation of mini-stumps, with successive collections of mini-cuttings every 30 days. N=36. Bars represented 5% confidence interval.

While several studies have investigated vegetative propagation with the mini-cutting technique for *P. reticulata*, to our knowledge, the literature lacks sufficiently robust and useful data that can be applied on a commercial scale. Rodrigues (2018) obtained 55.88% and 87.50% survival rates for apical and intermediate mini-cuttings, respectively, 45 days after staking. Without the application of plant growth regulators, Pessanha et al. (2018) reported that *P. reticulata* rooted 25.00, 0.01, and 8.33% in the collections carried out at 60, 90, and 150 days after apical pruning of the mini-stumps, respectively. These results are similar to our findings. On the other hand, *P. reticulata* rooted 30

and 31% when the whole leaf remained intact and when reduced by 75%, respectively (Neubert et al., 2017).

The process of callogenesis at the base of vegetative propagules may be an indicator of future rooting (Goulart et al., 2014). Once there is adventitious rooting, there are two patterns of root system emergence: the direct, in which rooting occurs with the emergence of the root directly from the propagule, and the indirect, in which the emergence of the root follows the formation of the callus (Xavier et al., 2013). In order for this to occur, the propagules must be kept longer under conditions adequate for root development (Oliveira et al., 2012; Pimentel et al., 2019). In this sense, Silva (2019) reported that *Paratecoma peroba* adventitious

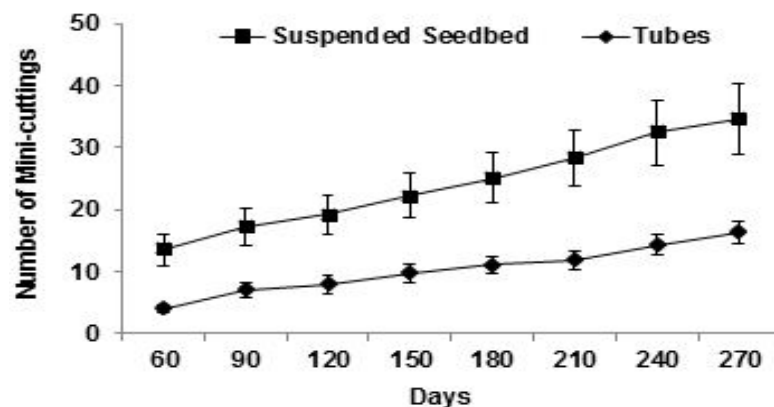


Fig. 3 Accumulated production of mini-cuttings with mini-stumps of *Plathymeria reticulata* Benth. carried out in two mini-gardens systems (suspended seedbed and tubes), 270 days after the formation of mini-stumps, with successive collections of mini-cuttings every 30 days. N = 36. Bars represent a 5% confidence interval..

Tab. 2 Evaluation of *Plathymeria reticulata* Benth. clonal seedlings, produced with the mini-cutting technique from propagules obtained in mini-stumps carried out in suspended seedbeds and tubes 120 days after staking.

Variables	Mini-garden		CV (%)
	Suspended Seedbed	Tubes	
H (cm)	8.87 b	11.86 a	46.79
D (mm)	2.69 a	1.87 b	31.68
LA (cm ²)	35.15 b	65.64 a	32.11
DMS (g)	0.39 a	0.48 a	55.25
DMSR (g)	0.10 a	0.13 a	82.88
NFOR	3.33 b	4.63 a	24.05
NRB	2.85 b	3.90 a	24.99
LFOR (cm)	9.74 a	8.97 a	23.36
TLR (cm)	318.74 a	464.71 a	29.35
DR (mm)	0.47 a	0.42 b	27.90
VR (cm ³)	0.54 a	0.68 a	64.81
DQI	0.053 a	0.056 a	74.60

rooting started after intense callus formation at the base of the mini-cuttings. The author found that 90% of the mini-cuttings had calluses, and roots were observed only 40 days after staking.

Callus formation at the base of mini-cuttings was also observed in other forest species. For *Aniba Rosaedora*, 26.66% of the vegetative propagules presented calluses (Menezes *et al.*, 2018). For *Ilex paraguariensis*, 44.14% and 47.72% of mini-cuttings presented calluses, without and with auxin application (2000 mg.L⁻¹ of IBA), respectively (Pimentel *et al.*, 2019).

Experiment 2 - Mini-garden management and clonal seedling production

The percentage of surviving *P. reticulata* mini-stumps was superior to that found in the literature for this species. 210 days after the establishment of a multiclonal mini-garden in tubes with seminal mini-stumps from 11 different matrices, Pessanha *et al.* (2018) found that only 8 progenies survived and the highest survival rate was 65%. For the same species, Neubert *et al.* (2017) observed mortality of mini-stumps beginning in the second collection of mini-cuttings. In the fourth and last collections, the highest percentage of mini-stump survival reported by the authors was 64.58%.

P. reticulata mini-stump survival rate variation among studies may be due to the genetic characteristics of the propagules, since neither nutritional nor water deficit issues of the mini-stumps were reported by the authors, nor were they observed in the present study during the course of the experiment.

On the other hand, the percentage of surviving *P. reticulata* mini-stumps was similar to that found for other native forest species, such as *Cedrela fissilis*, whose that the mini-garden had a survival rate of 100% after four months of sprout collection (Xavier *et al.*, 2003). Likewise, Oliveira *et al.* (2015b) observed a survival rate of 100% in *Handroanthus heptaphyllus* mini-stumps after 8 monthly collections.

The greater mini-cutting productivity when managed in suspended seedbeds might be explained by the area available for root growth. In contrast, irrigation and fertilization must be performed more often when using tubes due to their reduced volume (Silva *et al.*, 2012; Wendling *et al.*, 2005; Silva *et al.*, 2012; Souza *et al.*, 2015). This means that the lower the volume of the substrate, the higher the risk of nutritional deficiency, which can decrease mini-cuttings productivity.

When comparing two management systems in the formation of the clonal mini-garden (hydroponic system and in tubes) of *Erythrina falcata*, Cunha *et al.* (2008) found an average productivity of 2.9 mini-cuttings/mini-stump/collection for suspended seedbed and 1.3 for tubes (110 cm³) in four collections within 15 days. Oliveira *et al.* (2015), studying the mini-cuttings of *Handroanthus heptaphyllus* five months after the first pruning of mini-stumps managed in tubes (280 cm³), obtained a production of 1.47 mini-cuttings/mini-stump. For *Myracrodruon urundeuva* mini-stumps in plastic tubes with a capacity of 1,550 cm³, the average productivity was 1.7 mini-cuttings/mini-stump over nine collections with intervals of 28 days (Justino *et al.*, 2017). These findings indicate that the productivity of *P. reticulata* mini-stumps was similar to that observed for other native forest species.

Pessanha *et al.* (2018) used *P. reticulata* mini-cuttings 5 cm in length and the superior rate of cloned seedlings was 46.43%. Thus, the higher index of *P. reticulata* clonal seedling production (50%) found in the present study may be related to the size of the propagule, since shorter propagules may not have the necessary reserves for rooting and survival (Fachinello *et al.*, 2005; Braga *et al.*, 2006; Costa, Pinto and Bertolucci, 2007).

On the other hand, the larger the propagule, the greater the amount of carbohydrate reserves used as a carbon source for nucleic acid and protein biosynthesis (Fachinello *et al.*, 1995). However, very long propagules may be more susceptible to dehydration due to their larger exposed surface areas that therefore require larger quantities of water to supply the greater amount of living tissue (Lima *et al.*, 2006). However, Rodrigues (2018) used mini-cuttings of *P. reticulata* with 15 cm in length and observed lower percentages of propagule survival (13.33% for apical mini-cuttings, 18.33% for intermediate, and 16.67% for basal, N = 60).

Araújo *et al.* (2019), working with *Paratecoma peroba* mini-cuttings, found that 10 cm apical mini-cuttings showed the greatest survival (90%) and rooting (80%) rates, compared to shorter mini-cuttings, 90 days after staking. However, in the seedling production of *Azadirachta indica* by mini-cutting, Fernandes *et al.* (2017), using three sizes of mini-cuttings (5, 8, and 11 cm), observed that the size of the propagules did not affect rooting, although seedlings produced from mini-cuttings of greater length showed higher Dickson Quality Index (DQI) values.

The highest rooting percentages in propagules of greater lengths were also observed for *Spondias tuberosa*

(Rios et al., 2012), *Hibiscus rosa-sinensis* L. (Pizzatto et al., 2011), and *Ficus enormis* (Mireski et al., 2019) cuttings.

However, for vegetative propagules of *Piper umbellatum* L. (Gomes and Krinski, 2016) and *Ilex paraguariensis* (Pimentel et al., 2020), the size did not influence the survival or the rooting of the propagules. Thus, it is reasonable to conclude that the propagule size response with respect to rooting and survival varies according to the species.

Factors such as temperature and humidity, both in the rooting sector and in the greenhouse, can also affect the survival and rooting rates of mini-cuttings (Brondani, 2008; Xavier et al., 2013). According to reports in the literature concerning several forest species, while the ideal temperature for the rooting sector is between 20 and 30°C, the relative humidity should be at least 80% (Xavier et al., 2003; Brondani et al., 2007; Lamônica, 2013; Dias et al., 2015; Oliveira et al., 2016; Berude, 2018; Burin et al., 2018 a and; Rodrigues, 2018; Somavila, 2018; Souza, 2019). These values are similar to those observed in the present work and other works related to the vegetative propagation of *P. reticulata* (Neubert et al., 2017; Pessanha et al., 2018; Rodrigues, 2018). Therefore, temperature and humidity were not factors that led to the greater survival rate of the mini-cuttings in the present study.

The values regarding H, D, and DMS, observed in the present work are consistent with those found by Pessanha et al. (2018) for *P. reticulata* from the first sprout collection (60 days after the apical pruning of the mini-stump) five months after the staking of mini-cuttings. However, the values for these variables are lower than those the same authors obtained in later collections. In this way, there may be an improvement in the quality of the clonal seedlings in the present work with the use of propagules obtained during the execution of the mini-garden, as observed by Pessanha et al. (2018).

Differences observed between the number of first-order roots of the mini-cuttings between the mini-stumps management systems were also observed by Silva et al. (2012). Those authors found that *Toona ciliata* clonal seedlings originating from mini-cuttings from mini-stumps established in tubes (180 cm³) had a greater number of roots in the rooting sector. They hypothesized that the mini-cuttings from suspended seedbeds incurred higher energy costs for maintaining leaf turgor, resulting in reduced investment in adventitious rooting.

P. reticulata root numbers in mini-cuttings from both mini-garden management schemes were lower when compared to those reported by Pessanha et al. (2018), who studied the same species. However, those same authors performed assessment 150 days after staking. These values are also lower than those found for other tropical forest species, such as *Inga edulis*, which reached 9.3 and 13.7 roots without and with the application of growth regulators (AIB: 8000 mg.kg⁻¹), respectively, 45 days after staking (Berude et al., 2019), *Piptocarpha angustifolia* mini-cuttings, showing 6.3 roots/mini-cuttings, collected in winter (Ferriani et al., 2011), and *Cabralea canjerana* mini-cuttings, which presented averages of 2.20 to 6.46 roots/mini-cuttings at the same collection time 60 days after staking (Burin et al.,

2018). However, the values obtained for this variable were higher than those obtained for *Eugenia involucrata* mini-cuttings, which had 0.95 roots/mini-cuttings 120 days after staking (Chiele, 2016), and *Aniba Rosaedora* mini-cuttings, which presented 2.08 roots / mini-cuttings 90 days after staking (Menezes et al., 2018).

The Dickson Quality Index (DQI) values obtained for mini-cuttings collected from both mini-gardens was the same. They were lower than those found by Pessanha et al. (2018) while working with the same species (DQI at 150 days after staking: 3.99).

These results indicate that the mini-cutting technique is a viable alternative to seedling production with *P. reticulata*, especially when seed sourcing is a constraining factor in the production process. However, further studies aiming to increase rooting rates and test other management factors driving the rooting process are required.

CONCLUSION

The time in the mist chamber did not affect the rooting percentages of the mini-cuttings; 30 days remains the recommended period. While mini-stump root restriction reduced the production capacity of mini-cuttings, it did not affect the clonal seedling production index and the final quality of the clonal seedlings.

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Funding: DGB

Processing: GCMWC, DSP, RDS, MKFS

Analysis: GCMWC, DSP, RDS, MKFS

Writing: GCMWC, DSP, DGB

Review: GCMWC, DSP, DGB

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